

DITERPENES FROM THE RED ALGA *SPHAEROCOCCUS* *CORONOPIFOLIUS*

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Key Word Index—*Sphaerococcus coronopifolius*; Rhodophyta; diterpenes; structural determination.

Abstract—Four new diterpenes have been isolated from the red alga *Sphaerococcus coronopifolius*. Their structures have been established on the basis of chemical and spectral evidence.

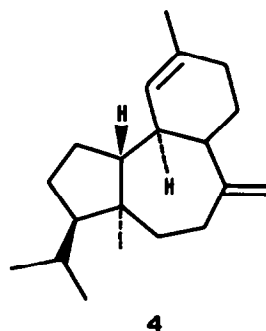
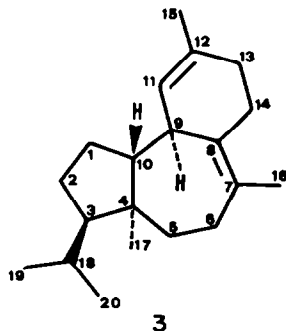
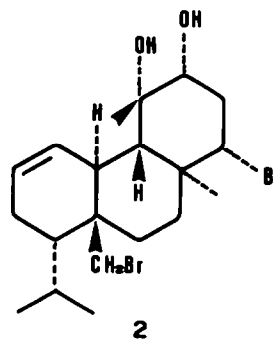
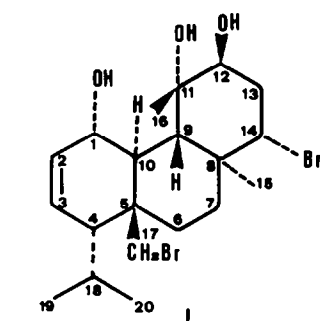
INTRODUCTION

Extensive studies in our laboratory on the constituents of the red alga *Sphaerococcus coronopifolius* have resulted in the isolation of a number of diterpenes which have been fully characterized and for which a general biogenetic scheme starting from geranylgeraniol has been proposed [1, 2].

Further studies on the same organism have now added four new compounds: 12*S*-hydroxybromosphaerodiol (1), 12*R*-hydroxybromosphaerol (2), isosphaerodiene-1 (3) and isosphaerodiene-2 (4), obtained in small amounts and identified on the basis of spectral and chemical evidence.

RESULTS AND DISCUSSION

The most polar component 1, $C_{20}H_{32}O_3Br_2$ (HRMS), contained three hydroxyl groups, as indicated by IR (3300–3050 cm^{-1}) and mass spectroscopy [M]⁺ m/z 482, 480, 478, [$M - H_2O$]⁺, [$M - 2H_2O$]⁺, [$M - 3H_2O$]⁺. On the basis of the 1H NMR spectrum two of the above groups had to be linked to secondary carbon atoms (δ 4.59, 1H, *dd* and 3.37 1H, *dd*). The 1H NMR spectrum also exhibited signals attributable to $-CH-CH=CH-CH-$ (δ 6.12, 1H, *dd* and 5.90, 1H, *dd*), bromomethine (δ 4.53, 1H, *dd*), bromomethylene (δ 3.55, 1H, *d* and 4.09, 1H, *d*), two secondary methyl groups (δ 1.03, 3H, *d* and 0.98, 3H,



d) and two tertiary methyl groups (δ 1.45, 3H, s and 1.27, 3H, s). Most structural detail was obtained from ^1H NMR analysis involving extensive double resonance experiments which indicated the partial structures (Table 1): $\text{C}_9\text{--}10\text{--C}_{11}\text{--}4\text{--C}_{18}\text{--}20$, $\text{C}_6\text{--}7$ and $\text{C}_{12}\text{--}14$.

The above data were interpreted in terms of the structure 1 based on a carbon skeleton present in several bromoditerpenes of *S. coronopifolius*. Confirmation of the proposed structure was obtained as follows.

12S-Hydroxybromosphaerol (5), a metabolite previously found in the same marine organism [3], by treatment with $\text{H}_2/\text{Pd-C}$ afforded the dihydroderivative 6 which was in turn obtained from 1 by catalytic hydrogenation in the presence of perchloric acid.

This result established the absolute configuration of 1, apart from the chirality of C-1, which must be *S* as indicated by the *J* value (10 Hz) between H-1 and H-10 indicative of the quasi-equatorial nature of the hydroxyl group.

The second compound (2) had very similar IR and MS spectra to those of 5. The ^1H NMR spectral characteristics of 2 were essentially identical to those of 5, except in the region influenced by the stereochemistry of C-12 (2: δ 2.36, 1H, *q*, *J* = 12 Hz, $\text{H}_{\text{ax}}\text{-}13$; 3.36, 1H, *dd*, *J* = 12 and 4 Hz, H-12; 3.90, 1H, *dd*, *J* = 12 and 3 Hz, H-14; 5: δ 2.72, 1H, *dt*, *J* = 3.5 and 12 Hz, $\text{H}_{\text{ax}}\text{-}13$; 3.47, 1H, *t*, *J* = 3.5 Hz, H-12; 4.48, 1H, *dd*, *J* = 3 and 13 Hz, H-14). The significant large differences in the chemical shifts and in the coupling constants of H-14, $\text{H}_{\text{ax}}\text{-}13$, and H-12 suggested that compounds 2 and 5 were C-12 epimers. This was confirmed by the conversion of 2 to sphaerococcenol (7), the major bromoditerpene from *S. coronopifolius* [4], by treatment with pyridinium chlorochromate in the presence of sodium acetate at room temperature.

Compound 3, $\text{C}_{20}\text{H}_{32}$, has not been previously reported as a naturally occurring compound, but has been synthesized by dehydration of presphaerol (8). It was

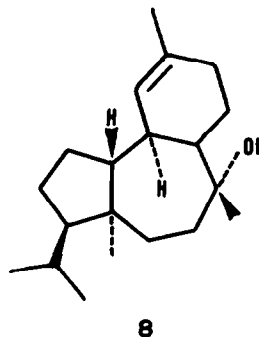
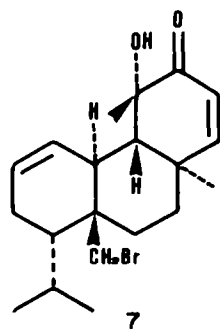
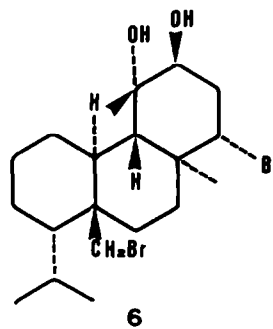
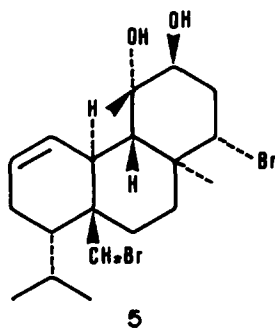
Table 1. ^1H NMR spectral data for compound 1

| H | δ (CDCl_3) | <i>J</i> (Hz) |
|-------|--------------------------------|-----------------|
| 1 | 4.59 (1H, <i>dd</i>) | 10 and 3 |
| 2 | 6.12 (1H, <i>dd</i>) | 10 and 3 |
| 3 | 5.90 (1H, <i>dd</i>) | 10 and 5 |
| 4 | 2.42 (1H, <i>dd</i>) | 6 and 5 |
| 6ax* | 1.26 (1H, <i>ddd</i>) | 14, 14 and 3 |
| 6eq† | 1.75–1.85 (2H, complex signal) | |
| 7ax* | | |
| 7eq† | 1.62 (1H, <i>ddd</i>) | 14, 4 and 3 |
| 9 | 2.01 (1H, <i>d</i>) | 10 |
| 10 | 2.59 (1H, <i>dd</i>) | 10 and 10 |
| 12 | 3.37 (1H, <i>dd</i>) | 3.5 and 3 |
| 13ax | 2.64 (1H, <i>ddd</i>) | 11, 11 and 3 |
| 13eq | 2.09 (1H, <i>ddd</i>) | 13, 3.5 and 3.5 |
| 14 | 4.53 (1H, <i>dd</i>) | 11 and 3.5 |
| 15 | 1.27 (3H, <i>s</i>) | |
| 16 | 1.45 (3H, <i>s</i>) | |
| 17 | 3.55 (1H, <i>d</i>) | 10.3 |
| 17' | 4.09 (1H, <i>d</i>) | 10.3 |
| 18 | 2.12 (1H, <i>m</i>) | |
| 19/20 | 0.98 (3H, <i>d</i>) | 7 |
| 20/19 | 1.03 (3H, <i>d</i>) | 7 |

*†Assignments may be reversed.

identified by comparison of its properties with those of an authentic sample [5].

The remaining compound (4) was an isomer of 3 and had very similar ^1H NMR spectral properties, the only difference being the absence of a vinylic methyl signal and the presence of two 1H broad singlets (δ 4.80 and 4.77) attributable to a $=\text{C}=\text{CH}_2$ group. Structure 4 was proved by careful analysis of the mixture obtained by dehydration



of presphaerol which revealed the presence of small quantities of 4.

EXPERIMENTAL

General. IR: CHCl_3 ; $^1\text{H NMR}$: 500 MHz, chemical shifts in ppm (δ) relative to TMS; prep. LC: Varian 5000 apparatus using a dual cell refractometer detector; optical rotations: CHCl_3 ; GC: coiled SE-30 fused silica capillary column (30 m \times 0.326 mm, J & W Scientific, Inc, H_2 as carrier gas, 140°).

Plant material. *Sphaerococcus coronopifolius* was collected at 8–12 m depth in the autumn of 1985 near Massalubrense, Bay of Naples. A voucher specimen is deposited at the Dipartimento di Chimica Organica e Biologica, Naples, Italy.

Extraction and purification. CHCl_3 extraction of the freeze-dried and ground alga (4 kg) afforded 15 g of a residue which was chromatographed on a silica gel (450 g) column. The polarity of the solvent (hexane) was increased with EtOAc until the solvent composition became hexane–EtOAc (3:2). Fractions exhibiting similar TLC profiles were combined to give fractions A, B and C in order of increasing polarity.

Fraction A (10 mg) was subjected to prep. HPLC (Si 60 LiChrosorb-Merck, eluent *n*-hexane) to give 3 (4.0 mg) and 4 (2.5 mg).

Compound 3, oily, $[\alpha]_D = +28^\circ$, had spectral (IR, MS and $^1\text{H NMR}$) and chromatographic (TLC, HPLC and GLC) properties identical with those of the product previously isolated by dehydration of presphaerol [5].

Compound 4, oily, $[\alpha]_D = +18^\circ$, IR $\nu_{\text{max}} \text{ cm}^{-1}$: 1650 and 895; $[\text{M}]^+$, m/z 272; $^1\text{H NMR}$ (CDCl_3): δ 0.84 (3H, *d*, $J = 7$ Hz, 3H-19/20), 0.94 (3H, *d*, $J = 7$ Hz, 3H-20/19), 1.66 (3H, *s* (*br*) 3H-15), 2.39 (1H, *m*, H-9), 4.80 and 4.77 (2H, *s* (*br*), *s*, 2H-16), 5.40 (1H, *d* (*br*), $J = 5$ Hz, H-11). Chromatographic (TLC, HPLC and GLC) and spectral (IR, MS and $^1\text{H NMR}$) properties of 4 were identical to those of an authentic sample synthesized as described below.

Fraction B (27 mg) was purified by HPLC (RP 18, LiChrosorb-Merck, eluent MeCN) to give 18 mg of pure 2, $[\alpha]_D = -34^\circ$; mp = 89–92°, IR $\nu_{\text{max}} \text{ cm}^{-1}$: 3350–3100; $[\text{M}]^+$, m/z 462, 464, 466; $^1\text{H NMR}$ (CDCl_3): δ 0.91 (3H, *d*, $J = 7$ Hz, 3H-19/20), 0.97 (3H, *d*, $J = 7$ Hz, 3H-20/19), 1.29 (3H, *s*, 3H-15), 1.46 (3H, *s*, 3H-16), 2.36 (1H, *q*, $J = 12$ Hz, H_{ax} -13), 3.04 (1H, *d* (*br*), $J = 6$ Hz, H-9), 3.36 (1H, *dd*, $J = 12$ and 4 Hz, H-12), 3.62 and 3.95 (1H each, AB system, $J = 10$ Hz, 2H-17), 3.90 (1H, *dd*, $J = 12$ and 3 Hz, H-14), 5.70 (1H, *m*, H-2), 6.04 (1H, *dd*, $J = 9$ and 2 Hz, H-1).

Fraction C (49 mg) was subjected to prep. HPLC (RP 18, LiChrosorb-Merck, eluent MeCN) to yield pure 1 (32 mg), $[\alpha]_D = 39^\circ$; mp = 88–90°; IR: 3300–3050 cm^{-1} ; HRMS: $[\text{M}]^+$ 478.07185 (calc. for $\text{C}_{20}\text{H}_{32}^{79}\text{Br}_2\text{O}_3$ 478.07191); $^1\text{H NMR}$: Table 1.

Dehydration of 8 to afford 3 and 4. CH_3COCl (0.5 ml) was added to a soln of 8 (42 mg) in xylene (2 ml) and the mixture was heated under reflux for 1 hr. After cooling, H_2O (2 ml) was added and the organic layer was taken to dryness. The residue was subjected to HPLC (RP 18, LiChrosorb-Merck, eluent MeOH) to give 3 (6 mg) and 4 (3 mg).

Oxidation of 2 to give 1. Pyridinium chlorochromate (13 mg) was added to a suspension of anhydrous NaOAc (4 mg) in CH_2Cl_2 (2 ml) containing 2 (14 mg). After stirring at room temp. for 2 hr the mixture was diluted with Et_2O and filtered. Evaporation of the filtrate and chromatography of the residue on TLC (silica gel) in CHCl_3 gave 6 mg of sphaerococcenol (7) [4].

Catalytic hydrogenation of 5 to give 6. 5 (23 mg) in EtOH (2 ml) was hydrogenated over 10% Pd/C (4 mg) at room temp. and atmospheric pressure for 3 hr. After removal of the catalyst by filtration, the soln was evaporated to dryness and the residue was purified by HPLC (RP 18, LiChrosorb-Merck, eluent MeOH) to give 7 mg of 6. $[\alpha]_D = -8^\circ$; mp = 95–97°; IR: 3300–3050 cm^{-1} ; $[\text{M}]^+$, m/z 464; $^1\text{H NMR}$ (CDCl_3): δ 1.02 (3H, *d*, $J = 7$ Hz, 3H-19/20), 1.07 (3H, *d*, $J = 7$ Hz, 3H-20/19), 1.29 (3H, *s*, 3H-16), 1.44 (3H, *s*, 3H-15), 1.78 (1H, *d*, $J = 13$ Hz, H-9), 2.15 (1H, *ddd*, $J = 13$, 4 and 4 Hz, H_{eq} -13), 2.46 (1H, *ddd*, $J = 13$, 13 and 3 Hz, H-10), 2.73 (1H, *ddd*, $J = 13$, 13 and 4 Hz, H_{ax} -13), 3.41 (1H, *t*, $J = 4$ Hz, H-12), 3.92 (2H, AB system, $J = 9.5$ Hz, 2H-17), 4.48 (1H, *dd*, $J = 13$ and 4 Hz, H-14).

Reduction of 1 to obtain 6. A soln of 1 (7 mg) in HOAc (1 ml) and perchloric acid (0.1 ml) was hydrogenated over 10% Pd/C (3 mg) for 2 hr at room temp. and atmospheric pressure. Following the usual work up compound 6 (3 mg) was isolated by HPLC.

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